



**Meise
Botanic Garden**

Postprint

This is the accepted version of a paper published in *Plant Biology*. This paper has been peer-reviewed but does not include the final publisher proof-corrections or journal pagination. The definitive version is available at <https://doi.org/10.1111/plb.13631>. Access to the published version may require subscription. When citing this work, please cite the original published paper (citation below).

Please cite this article as:

Van Rossum, F. and Le Pajolec, S. (2024), Maternal effects and inbreeding depression in post-translocation progeny of *Campanula glomerata*. *Plant Biol J*, 26: 427-436.
<https://doi.org/10.1111/plb.13631>.

1 **Maternal effects and inbreeding depression in post-translocation progeny of *Campanula***
2 ***glomerata***

3
4

5 Fabienne Van Rossum^{1,2}, Sarah Le Pajolec¹

6

7 ¹Meise Botanic Garden, Nieuwelaan 38, 1860 Meise, Belgium

8 ²Service général de l'Enseignement supérieur et de la Recherche scientifique, Fédération
9 Wallonie-Bruxelles, rue A. Lavallée 1, 1080 Brussels, Belgium

10

11

12 **Running head:** Maternal and inbreeding effects in post-translocation progeny

13

14

15 Corresponding author: Fabienne Van Rossum; Meise Botanic Garden, Nieuwelaan 38, 1860
16 Meise, Belgium; e-mail: fabienne.vanrossum@botanicgardenmeise.be; tel: +32 2 2600932

17

18

19 **Keywords:** inbreeding depression; maternal genetic and environmental effects; mixed seed
20 sources; phenotypic plasticity; plant translocations

21

22

23 **Key message of the paper:**

24 Inbreeding depression and strong maternal effects were detected in post-translocation progeny
25 fitness related to maternal transplant seed source origin and lineage in translocated populations
26 of *Campanula glomerata*, which might affect global genetic diversity patterns in the long term.

27

28

29

30 **Abstract**

- 31 • Evaluation of plant translocation success based on fitness-related quantitative traits
32 combined with molecular markers may contribute to a finer assessment of inbreeding,
33 selective and rescue processes, which may have long-term consequences for population
34 dynamics and viability.
- 35 • We investigated fitness traits (seed germination, seedling viability, and juvenile growth and
36 mortality) combined with 15 microsatellite loci of the first post-translocation seed progeny
37 from two translocated populations of *Campanula glomerata*, an insect-pollinated, self-
38 incompatible perennial herb. We examined whether inbreeding, heterosis through admixture,
39 translocation site and maternal transplant seed source origin and lineage might affect seed
40 quality and juvenile growth in controlled cultivation conditions.
- 41 • Flower production and seed germination of the transplants was higher in one of the two
42 translocation sites, which might be related to differences in soil and vegetation composition
43 and cover. Strong maternal effects related to seed source origin and lineage were found on
44 progeny size, with the largest transplants producing the largest progenies. The differences in
45 rosette diameter were maintained across the whole growth period measured. There was
46 inbreeding depression (rather than heterosis) related to biparental inbreeding at early progeny
47 growth stage, also expressed through juvenile mortality.
- 48 • Our findings highlighted that maternal transplant origin, especially when seed sources
49 consisted of small fragmented remnants, might have a selective value on fitness in the post-
50 translocation generations. If maternal effects and inbreeding depression persist, they might
51 affect global genetic diversity patterns in the long term. Further admixture in the next
52 generations might buffer maternal and inbreeding effects or lead to outbreeding depression.

53
54
55

ACCEPTED

56 INTRODUCTION

57 Reintroductions in extirpated populations and reinforcements of inbred and genetically
58 impoverished populations through plant translocations has become a popular restoration practice
59 for the recovery of viable and evolutionarily resilient populations of critically endangered
60 species (e.g., Zimmer *et al.* 2019; Fenu *et al.* 2023). Genetic monitoring based on neutral
61 molecular markers (microsatellites, SNPs) can evaluate whether genetic restoration has been
62 effective in reinforced populations and whether translocation has recreated populations with a
63 highly diverse gene pool (e.g., Van Rossum *et al.* 2020; Van Rossum & Hardy 2022 and
64 references cited therein; Dillon *et al.* 2023). For instance, a successful increase in genetic
65 diversity has been found in genetically depauperate populations after reinforcement by sowing
66 seeds or planting young plants of local or non-local provenance, for *Arnica montana*, an insect-
67 pollinated self-incompatible clonal herb (Van Rossum *et al.* 2020), and for *Pulsatilla vulgaris*,
68 an insect-pollinated predominantly outcrossing herb (Gargiulo *et al.* 2019). Using multiple seed
69 sources in mixture for plant translocation could provide new populations with high genetic
70 diversity and genetic composition representative of the wild genetic pool, as reported for several
71 insect-pollinated herb species, such as the self-compatible *Arenaria grandiflora* (Zavodna *et al.*
72 2015) and *Dianthus deltoides* (Van Rossum & Le Pajolec 2021), and the self-incompatible
73 *Campanula glomerata* (Van Rossum *et al.* 2023a), and for bird-, mammal- and/or insect-
74 pollinated self-compatible small tree or shrub species, such as *Banksia brownii* (Dillon *et al.*
75 2023) and *Lambertia orbifolia* (Monks *et al.* 2021). However, new cohorts in translocated
76 populations of *B. brownii*, *D. deltoides* and *L. orbifolia* were characterized by high selfing rates,
77 while restricted seed and pollen dispersal distances, already from the first recruited generation,
78 were found in translocated populations of *C. glomerata*. Those mating patterns might be related
79 to insufficient availability of dispersal vectors, pollinator behaviour related to local floral
80 abundance and plant density, and/or translocation site environment (Monks *et al.* 2021; Dillon
81 *et al.* 2023; Van Rossum 2023; Van Rossum *et al.* 2023a). When seed source populations are of
82 small size or with high clonal extent, only a small number of potential parents may contribute to
83 the seeds used for propagating transplants. As a result, many transplants can be related (i.e. full
84 sibs or half sibs), so that biparental inbreeding may also arise in the newly established cohorts
85 (Aguilar *et al.* 2019; Doyle *et al.* 2023), as reported for *D. deltoides* (Van Rossum & Le Pajolec
86 2021), for *Primula vulgaris*, a wild bee-pollinated heterostylous herb (Barmantlo *et al.* 2018),
87 and for *Hakea nitida*, a bird- and insect-pollinated shrub (Millar *et al.* 2021).

88
89 Small populations may show inbreeding depression, i.e. reduced fitness of inbred seed progeny,
90 but also drift load, that is reduced fitness of outbred offspring due to the accumulation and
91 fixation of recessive deleterious mutations over time through genetic drift (Paland & Schmid
92 2003; Willi *et al.* 2005). In the case of reinforcement of declining populations suffering from
93 inbreeding depression and/or drift load, whether there is genetic rescue has to be evaluated, so
94 whether introducing new genetic variation and/or heterosis (higher fitness of cross progeny)
95 have counteracted inbreeding and drift issues (Bell *et al.* 2019). Genetic rescue cannot be
96 evaluated based on neutral molecular markers only. Fitness-related quantitative traits are
97 required (Willi *et al.* 2006; Nicotra *et al.* 2015). Moreover, inbreeding (or outbreeding)
98 depression, and local adaptation issues, with maladaptation possibly related to maternal effects
99 in case of differences between source and translocation environments, may arise in translocated
100 populations (Vergeer *et al.* 2004; Edmands 2007; Schuler & Orrock 2012; Zavodna *et al.* 2015).
101 For instance, despite evidence from neutral molecular markers of global increase in genetic
102 diversity and of genetic mixing (admixture) in the newly established post-translocation
103 generations, reduced plant size, floral display or reproductive performance in inbred offspring,
104 indicating inbreeding depression, was found in translocated populations of *B. brownii* (Rodger *et al.*
105 *et al.* 2023), *D. deltoides* (Van Rossum & Le Pajolec 2021), *A. grandiflora* (Zavodna *et al.* 2015),

106 and *P. vulgaris* (Barmantlo *et al.* 2018). For the last two species, reduced fitness in admixed
107 offspring (so with parents from different source origins), indicating outbreeding depression and
108 local adaptation as well as heterosis for within-population outcrossed offspring, were detected.
109 Maternal effects at early growth progeny stages as well as high phenotypic plasticity at later
110 stages was reported for *A. montana* (Van Rossum *et al.* 2020) and *D. deltooides* (Van Rossum &
111 Le Pajolec 2021). Local adaptation through increased biomass and flowering of local transplants
112 has been reported in *Succisa pratensis*, an insect-pollinated self-compatible herb (Vergeer *et al.*
113 2004). Maladaptation of non-local transplants indicated by lower reproductive success was
114 found for *Silene douglasii* (Lofflin & Kephart 2005). Therefore, investigating fitness-related
115 quantitative traits combined with molecular markers may contribute to a finer assessment of
116 inbreeding, selective or rescue processes occurring in translocated populations, which may have
117 long-term consequences for population dynamics and viability. It can provide information for
118 proposing adaptive-oriented management of the populations (Willi *et al.* 2006; Van Rossum &
119 Hardy 2022; Dillon *et al.* 2023; Rodger *et al.* 2023).

120
121 In this paper, we investigated fitness-related quantitative traits (seed germination, seedling
122 viability, juvenile growth and mortality) combined with molecular markers (microsatellites) of
123 the first post-translocation seed progeny produced by the transplants, in translocated populations
124 of the insect-pollinated, self-incompatible, *Campanula glomerata* L. (Campanulaceae). Local
125 small populations were used as seed sources for plant translocations in southern Belgium
126 (Godefroid *et al.* 2016). Previous genetic studies (Van Rossum *et al.* 2022, 2023a) based on
127 neutral nuclear microsatellite markers showed that the adult generation of these small
128 populations still retained high, likely old (pre-fragmentation) genetic diversity. For the offspring
129 generation (used for transplant propagation in controlled cultivation conditions), there was
130 evidence of extensive contemporary pollen dispersal within populations. However, demographic
131 bottlenecks, genetic erosion (due to genetic drift effects) and biparental inbreeding were found in
132 some of the source populations, as well as inbreeding depression, and differences in plant size
133 among seed source populations (Van Rossum *et al.* 2022). Given the small effective population
134 sizes (Van Rossum *et al.* 2022), drift load might also be expected (Paland & Schmid 2003; Willi
135 *et al.* 2005, 2013). In the translocated populations, genetic admixture between seed source
136 origins was favoured by the mixed spatial arrangement of the transplants, but seed and pollen
137 dispersal at short distances has led to spatial structuring of the genetic variation of the newly
138 established recruits (Van Rossum *et al.* 2023a). Mating among relatives might be expected in the
139 subsequent post-translocation generations, which might lead to biparental inbreeding but also to
140 reduced seed set (Aguilar *et al.* 2019; Doyle *et al.* 2023). Therefore, examining whether
141 inbreeding depression and heterosis in admixed offspring (indicating drift load; Willi *et al.* 2013)
142 might occur, and whether translocation site environment, and maternal transplant seed source
143 origin and lineage (and so maternal genetic and environmental effects), might affect seed quality
144 and progeny growth of the first post-translocation generation, can provide some additional
145 insights on possible outcomes for the next generations. More precisely, we addressed the
146 following questions: (1) Does transplant seed quality (seed germination, seedling viability, and
147 juvenile mortality) and seed progeny fitness (rosette diameter and number of leaves) differ
148 between translocation sites, reflecting environmental effects, and thus possible future selective
149 processes? (2) Does progeny fitness differ among maternal transplant seed source origins and
150 transplant site lineages, reflecting possible maternal genetic and environmental effects, and are
151 the differences in progeny fitness maintained with time or is there evidence of phenotypic
152 plasticity? (3) Is there evidence of (biparental) inbreeding / outbreeding depression (lower fitness
153 of inbred / admixed offspring) and of heterosis (higher fitness of the between-source progeny) in
154 the seed progeny?

155 MATERIALS AND METHODS

156 Study species

157 *Campanula glomerata* L. (Campanulaceae) has a wide continental Eurasian distribution range,
158 and typically occurs in calcareous grasslands, scrub, open woodlands and sand dunes (habitat
159 code 6210 according to EU 92/42/EEC Habitats Directive) (Klotz *et al.* 2002; Denisow &
160 Wrzesień 2015). Those habitats, and their associated species, have strongly declined in
161 northwestern Europe, with plant populations becoming small and fragmented (Hill *et al.* 2004;
162 Bachmann & Hensen 2007; Godefroid *et al.* 2016). In Belgium, *C. glomerata* is critically
163 endangered, with a few small remaining wild populations, occurring in degraded calcareous
164 habitats (evolving into more competing, nutrient-richer hay meadow vegetation) (Godefroid *et*
165 *al.* 2016; Godefroid, unpublished data).

166
167 This herbaceous perennial species is characterized by a gametophytic self-incompatible breeding
168 system, and can be considered as an obligate outcrosser (Gadella 1964). Plants flower in June-
169 August (Klotz *et al.* 2002; Bachmann & Hensen 2007). Flowers produce nectar and are
170 pollinated by insects, mainly bumble bees, solitary bees, honey bees and hoverflies (Albrecht *et*
171 *al.* 2007; Denisow & Wrzesień 2015). The fruit is a capsule containing numerous and tiny seeds
172 (up to 100), which are dispersed over short distances by capsule shaking but also possibly over
173 longer distances through zoochory by grazing sheep. Seeds do not form a persistent seed bank in
174 the soil, but plants can live for 25-30 years and subsist as non-sprouting rhizomes (Klotz *et al.*
175 2002; Bachmann & Hensen 2006, 2007).

176 177 Study populations and sampling

178 Plant translocations (500 transplants in each site propagated from mixed seed sources) were
179 conducted in four sites (restored calcareous grasslands, planting on scraped, bare soil) from
180 Belgian Lorraine (southern Belgium) in September 2015 (Fig. 1; Godefroid *et al.* 2016; Van
181 Rossum *et al.* 2022). Fitness measures and seed collection for the present study were conducted
182 in two translocated populations, Ficherulle (FICH; 49°42'54"N, 5°13'31"E) and Sainte-Cécile2
183 (CEC2; 49°43'40"N, 5°14'00"E), located at 1.53 km from each other. The two translocation
184 sites differed in soil and vegetation cover and composition: FICH was characterized by a sandy-
185 marly soil and a dense (80% cover on average) grassland vegetation (dominated by tall herb
186 species, in particular *Tanacetum vulgare* L., *Leucanthemum ircutianum* DC., *Rubus* sp.,
187 *Rhinanthus angustifolius* C.C. Gmel., *Silene latifolia* Poir., *Centaurea scabiosa* L. and *Dactylis*
188 *glomerata* L.), and CEC2 by a marly soil and sparse (50% cover on average) grassland
189 vegetation (dominated by *Holcus lanatus* L., *Bromopsis erecta* (Huds.) Fourr. and by short forb
190 species, in particular *Ranunculus repens* L., *Trifolium repens* L., *Taraxacum* F.H. Wigg., and
191 *Geranium columbinum* L.) (Fig. S1).

192
193 The transplants of the translocated populations originated from seeds collected in five small wild
194 populations (flowering population size ranging 7-28 flowering individuals): Chassepierre
195 (CHA), Emond (EMO), Fontenoille (FON), Lambermont (LAM) and Watrinsart (WAT) (Van
196 Rossum *et al.* 2022, 2023a). Based on sibship assignment analyses (Van Rossum *et al.* 2022),
197 but considering *C. glomerata* as self-compatible in the parameters, no evidence of breakdown of
198 the self-incompatibility system leading to selfing was found in the small seed source populations
199 (Van Rossum, unpublished results). The percentage of offspring (used as transplants for plant
200 translocation) sharing at least one parent (full sibs and half sibs) ranged 9.0-54.6% (Figure 1),
201 and biparental inbreeding was detected in CHA and LAM (significantly positive F_{IS} values after
202 correction for null allele frequency = 0.016 and 0.043, respectively) (Van Rossum *et al.* 2022).

203

204 Seeds (F1 generation) were collected in 2016 on 188 transplants (95 and 93 in CEC2 and FICH
205 translocation sites, respectively, and 16 to 20 per seed source origin; Fig. 1). They were stored in
206 a dry-cold environment (15°C, 15% relative moisture), until the germination experiment.
207

208 Maternal effects were taken into account by seed source origin of the maternal transplants, by
209 maternal lineage, and by two quantitative variables. First, the number of flowering stems and of
210 flowers per stem (for max. five flowering stems) was counted in 2016 for the 188 transplants,
211 and used to estimate floral display size as the total number of flowers per transplant (Godefroid
212 & Van Rossum, unpubl. data). Second, rosette diameter (in cm) measured 2 months after
213 germination (just before translocation) was used as an estimator of vegetative maternal
214 transplant size (data from Le Pajolec *et al.* 2021).
215

216 **Quantitative trait measures of seed progeny**

217 At the beginning of April 2017 (after 1 month at 5°C to break the dormancy), 100 seeds for each
218 of the 188 transplants were germinated in Petri dishes on 1% agar (10 g/L) in incubators (20°C,
219 8/16 light/dark photoperiods). Seed germination (the proportion of germinated seeds) and the
220 proportion of viable seedlings (i.e. fully developed green seedlings over the total -chlorotic and
221 green- number of seedlings, excluding rotten seedlings) were recorded after three weeks. A total
222 of 1,215 viable seedlings (up to 12 per transplant, for 146 maternal transplants) were transferred
223 to an unheated greenhouse in pots of 3 cm diameter, and repotted four weeks later in 9 cm pots
224 with (pH 6.5) extra fine potting soil (Godefroid *et al.* 2016). The pots were randomized every
225 two weeks. When the plants were 6-weeks old, we started to measure plant size, i.e. rosette
226 diameter (in cm) every two weeks (measurement repeated four times). The number of leaves was
227 counted at the end of the experiment (12 weeks after germination). We also recorded mortality
228 (the proportion of dead plants) at the end of the experiment (for fitness datasets, see Van Rossum
229 *et al.* 2023b). The 12 week-old plants were delivered to practitioners (Natagora) for translocation
230 in two additional sites in Belgian Lorraine.
231

232 **Individual inbreeding and parental seed source origin assessed from multilocus genotypic 233 data**

234 In order to test for heterosis (higher fitness of the heterozygotes) and (biparental) inbreeding /
235 outbreeding depression (lower fitness of inbred / admixed offspring) in the seed progeny, fitness
236 quantitative traits were analysed in relation to multilocus genotypic data based on 15
237 microsatellite loci obtained in a previous study (Van Rossum *et al.* 2023a) for 166 and 136
238 transplants and for 122 and 125 seed progenies (1-3 offspring per maternal transplant) in CEC2
239 and FICH translocation sites, respectively (for datasets, see Van Rossum & Godé 2021, 2023).
240 For this purpose, Homozygosity by Loci (HL), as an indicator of individual inbreeding level,
241 was computed for each sampled transplant and seed progeny, separately in each population,
242 using the software GENHET (Coulon 2010). HL can vary from 0 (all loci are heterozygous) to 1
243 (all loci are homozygous; Aparicio *et al.* 2006). One microsatellite locus (CAM27) was excluded
244 from the analyses, because its high frequency of null allele (ranging 28-38%; Van Rossum &
245 Godé 2022) might falsely inflate HL.
246

247 The five seed source populations were found to be genetically differentiated from each other
248 based on 15 microsatellite loci (F_{ST} values between populations ranging 0.089-0.159), and
249 corresponded to distinct clusters in STRUCTURE (Pritchard *et al.* 2000) Bayesian clustering
250 analysis (Van Rossum *et al.* 2023a). Transplants could be assigned to their seed source origin
251 (CHA, EMO, FON, LAM or WAT) with high cluster membership values ($Q \geq 80\%$). Therefore,
252 it was possible to identify the seed source origin of the parental transplants of the seed progeny
253 from the STRUCTURE analysis, and so whether both parents belonged to the same or different

254 source origins (Van Rossum *et al.* 2023a). Two cross categories (within or between seed
255 sources) were therefore defined based on cluster membership (Q) values inferred from
256 STRUCTURE analysis, i.e. whether seed progeny descended from natural crosses between
257 parental transplants of the same or different source origins, respectively. The threshold was set at
258 $Q = 80\%$, with within-source crosses assumed when maximum Q value for the best cluster was \geq
259 80% and between-source crosses (admixed individuals) when Q value was $< 80\%$ in all clusters
260 (Van Rossum *et al.* 2023a).

261

262 **Data analysis**

263 We conducted two-way analyses of covariance (ANCOVA) together with pairwise Tukey HSD
264 post-hoc tests for testing for differences in fitness variables in relation to translocation site, seed
265 source origin and lineage of the maternal transplants and/or maternal (transplant) fitness (Table
266 S1, Fig. 1). We used a Generalized Linear Model (logit function) for the total number of flowers
267 (negative binomial distribution) and for proportion variables (seed germination, viable seedlings,
268 juvenile mortality; binomial distribution, taking overdispersion into account by dividing the
269 deviance by its degrees of freedom to estimate the dispersion parameter; McCullagh & Nelder
270 1989). The transplant variables were tested in relation to translocation site and seed source origin
271 of the maternal transplants, with rosette diameter, total number of flowers (log-transformed) and
272 HL (Box-Cox-transformed) of the maternal transplants as covariates (Table S1). Differences in
273 transplant HL between the five source origins were tested by performing an one-way Analysis of
274 Variance (ANOVA) together with pairwise Tukey tests.

275

276 For seed progeny, leaf number (Box-Cox-transformed) and rosette diameter (as repeated
277 measures over time: t1 to t4; with Greenhouse-Geisser adjusted probabilities) were tested using a
278 linear model in relation to translocation site, seed source origin of the maternal transplants, and
279 maternal transplant lineage (nested in source origin), with rosette diameter, total number of
280 flowers and HL of the maternal transplants as covariates (Table S1). The effects of categorical
281 variables and covariates were tested against the variation among maternal transplant lineages.
282 The effect of maternal transplant lineage was tested against the residual (error) variation among
283 individuals. The analyses were also conducted on the data subset analysed with molecular
284 markers, in relation to translocation site, seed source origin of the maternal transplants, and
285 progeny HL as covariate. As only 31 out of 247 seed progenies were identified as within-source
286 cross category, Mann-Whitney U tests were performed for testing for differences in rosette
287 diameter, leaf number and progeny HL between the two cross categories. We performed
288 Pearson's or Gamma correlation analyses (for normally- and non-normally-distributed variables,
289 respectively) between progeny or transplants fitness variables and HL. All analyses were
290 performed using STATISTICA version 12 (Dell Inc., Tulsa, Oklahoma, USA).

291

292 **RESULTS**

293 **Translocation site environmental effect**

294 There was a translocation site effect on maternal transplant fitness variables (Table S2): the total
295 number of flowers and seed germination were significantly higher in CEC2 than in FICH
296 translocation site (Fig. 2ab, Table S3). Seed germination was low (≤ 0.49 at individual maternal
297 transplant level; Fig. 2), but most seedlings were viable (Table S3), as only 22 out of the 2,038
298 germinated seedlings were chlorotic (12 in CEC2 and 10 in FICH translocation site). Therefore,
299 the proportion of viable seedlings was not further considered in the analyses. There was no effect
300 of translocation site on juvenile mortality and on seed progeny fitness variables (Fig. 2cf and S2,
301 Tables S2, S5 and S6).

302

303 **Maternal seed source origin and lineage effects**

304 There was a seed source origin effect on maternal transplant fitness variables. The transplants of
305 CHA seed source origin produced on average more flowers than the transplants originating from
306 EMO, FON and WAT (Tukey tests, $P = 0.003-0.014$; Fig. 2a, Tables S2 and S3). Juvenile
307 mortality was higher for maternal transplants originating from FON than from LAM and from
308 WAT (Tukey tests, $P = 0.020$ and 0.006 , respectively; Tables S2 and S3; Fig. 2c). The other
309 pairwise Tukey tests testing for differences in total number of flowers, seed germination or
310 juvenile mortality between seed source origins were not significant (P ranging from 0.059 to
311 0.998).

312
313 For seed progeny, we found a significant effect of seed source origin of the maternal transplants
314 on progeny rosette diameter for the four measured periods of growth, with WAT and LAM seed
315 source origins growing larger seed progeny plants (Fig. 2de and S2, Tables S4 and S5). Progeny
316 of WAT transplants produced less leaves than those of CHA and EMO, but only in CEC2
317 translocation site (Fig. 2f). We found significant differences in progeny plant size (rosette
318 diameter, leaf number) among maternal transplants (Tables S5 and S6). Progeny rosette diameter
319 increased with maternal rosette diameter at t1, t2 and t3 (Pearson's correlation coefficient
320 between maternal rosette diameter and average progeny rosette diameter per maternal transplant
321 $r = 0.319, 0.309$, and 0.245 , respectively, $P \leq 0.003$; Fig. 3), while at t4 the positive correlation
322 was only a trend ($r = 0.155, P = 0.064$). Progeny rosette diameter and leaf number were not
323 significantly ($P > 0.05$) related to maternal floral display size, except at t4 (β estimate of the
324 ANCOVA = $-0.176, P = 0.022$, but Pearson's correlation coefficient between maternal floral
325 display size and average progeny rosette diameter per maternal transplant $r = -0.080, P = 0.338$)
326 and to maternal HL (Tables S5 and S6). Maternal rosette diameter maternal and floral display
327 size were not correlated ($r = 0.086, P = 0.239$).

328 329 **Testing for biparental inbreeding / outbreeding depression and heterosis**

330 There was no significant correlation between maternal HL and the total number of flowers of the
331 transplants (Pearson's correlation coefficient $r = 0.082, P = 0.315$), seed germination and
332 juvenile mortality (Gamma correlation coefficient $\Gamma = 0.061$ and $0.031, P = 0.271$ and 0.613 ,
333 respectively). The ANOVA on maternal HL showed a significant effect of seed source origin
334 ($F_{(4,148)} = 3.04, P = 0.019$); this was only due to significantly higher HL values for transplants
335 originating from FON than for WAT seed source (mean value \pm SE = 0.26 ± 0.03 and $0.15 \pm$
336 0.02 , respectively; Tukey test, $P = 0.010$). The other pairwise Tukey tests testing for differences
337 in transplant HL between seed source origins were not significant ($P = 0.112-0.994$).

338
339 Progeny HL was negatively correlated with progeny rosette diameter at the first measure (t1) (Γ
340 = $-0.090, P = 0.038$), but it was not correlated with older stages of rosette diameter and with leaf
341 number (Γ ranging from -0.075 to $0.012, P \geq 0.082$). Rosette diameter at t1 was significantly
342 higher by 10% on average for between-source cross category than for within-source cross
343 category (mean value \pm SE = 11.3 ± 0.2 cm and 10.0 ± 0.5 cm, respectively; Mann-Whitney U
344 tests, $Z = 2.30, P = 0.021$; Fig. 4, Table S4), but the differences were not significant neither for
345 the next measures of rosette diameter (Z values ranging with time from 0.80 to $0.67, P > 0.421$)
346 nor for leaf number ($Z = -0.31, P = 0.754$). Progeny HL was significantly higher for within-
347 source cross category than for between-source category (HL ranging from 0.060 to 0.505 and
348 from 0 to 0.365 , respectively; $Z = -4.79, P < 0.001$). When performing a Mann-Whitney U test
349 for the same range of progeny HL (HL = $0.060-0.302$), the test was not significant anymore ($Z =$
350 $1.25, P = 0.210$), indicating that the difference in rosette diameter at t1 could be rather ascribed
351 to HL (inbreeding depression) than to a between-source cross effect (heterosis).

352 353 **DISCUSSION**

354 **Differences in transplant reproductive fitness between translocation sites**

355 Seed germination in controlled conditions (after three weeks) was low, but similar to values
356 reported in other studies (Bachmann *et al.* 2005; Koutsovoulou *et al.* 2014). The high number of
357 seeds per capsule (up to 100 seeds; Bachmann & Hansen 2007) and per plant (94.5 and 48.9
358 flowers on average in CEC2 and FICH translocation sites, respectively; Table S3) can however
359 compensate for this low germination rate. Reproductive fitness of the transplants was higher in
360 CEC2 than in FICH translocation site, with transplants producing more flowers and higher seed
361 germination, and so possibly better seed quality, despite similar global genetic diversity (based
362 on neutral molecular markers) and inbreeding levels between sites (Van Rossum *et al.* 2023a).
363 The two translocation sites differ in environmental conditions, in particular in soil and vegetation
364 cover and composition, with a more sandy (vs. a more marly) soil and a denser vegetation in
365 FICH than in CEC2 (Fig. S1), so that there may be differences in resource and water availability,
366 shading and competition, but also in pollinator services between sites. Differences in abundance
367 and composition of simultaneously flowering nectar-producing species might increase
368 competition or facilitation (through attractiveness) for pollinators, reducing or increasing pollen
369 transfer efficiency, which may in turn affect seed quality (Mitchell *et al.* 2009). Flowering of *C.*
370 *glomerata* has been reported to be reduced by shading from neighbour plants of competing
371 species (Bachmann *et al.* 2005), and seed quality in *C. glomerata* was found to be related to
372 pollination services, through seed mass increasing with pollinator species richness and
373 abundance (Albrecht *et al.* 2007). Increasing maternal nutrient resources and light intensity were
374 associated with increasing seed mass and decreasing seed germination in *Campanula americana*
375 (Galloway 2001, 2005).

376
377 **Differences in seed progeny growth according to maternal transplant seed source origin**
378 **and lineage**

379 No effect of translocation site origin was detected on further life stages, but there were
380 differences in progeny growth according to maternal transplant seed source origin and lineage.
381 These differences in progeny growth were coherent with differences in plant size found among
382 transplants before planting (Van Rossum *et al.* 2022): large transplants, one year after
383 translocation, also produced large progenies in controlled cultivation conditions (Fig. 3),
384 suggesting that there might be a genetic basis of the maternal effects on early growth stage
385 (Roach & Wulff 1987). Moreover, the differences in rosette diameter found in controlled
386 conditions between some maternal transplant seed source origins and lineages were maintained
387 across the whole growth period measured, suggesting strong maternal genetic and/or
388 environmental effects. Strong maternal environmental effects can be expected in the case of very
389 restricted seed dispersal (as found for *C. glomerata*), as seeds will germinate at proximity of the
390 maternal plants, and so in the mother's environment (Galloway 2005). Our findings on *C.*
391 *glomerata* contrast thus with similar experimental studies on *A. montana* and *D. deltoides*, which
392 had found evidence of considerable phenotypic plasticity. Indeed, for these species, differences
393 in plant size of the F1 generation found during the first weeks of growth had already disappeared
394 after eight weeks of cultivation (Van Rossum *et al.* 2020; Van Rossum & Le Pajolec 2021).
395 Progeny rosette diameter at t4 tended to decrease with increasing maternal floral display size.
396 However, the correlation was not significant with average values, suggesting that this finding
397 was not indicative of maternal effects, but rather reflected processes related to pollination and to
398 resource limitation. For instance, competition for resources among fruits or developing seeds,
399 possibly combined with pollination (and so seed development) timing, might affect individual
400 seed quality, and consequently offspring size, and might be higher in large floral displays
401 (Fenner & Thompson 2005).

402

403 Phenotypic plasticity can provide ability to adapt to changing or new environments, while strong
404 maternal environmental and genetic effects can reflect local adaptation (Galloway 2005; Schuler
405 & Orrock 2012; Nicotra *et al.* 2015). If this large range of variation among maternal lineages
406 suggests high –possibly adaptive– genetic variability in the translocated populations (Hamilton
407 *et al.* 2017), we may wonder whether such fitness differences might be maintained or
408 exacerbated under field conditions (expected to be more stressful –especially drier– than
409 cultivation conditions). The maintenance of fitness differences might result in differential
410 transplant survival and reproductive success in translocation sites. In particular, the adults of
411 small populations used as seed sources might consist of old, resilient genotypes, which had
412 survived in unfavourable conditions, in particular, degraded habitat quality due to grassland
413 recolonisation by dense, competing, cover vegetation resulting in nutrient-richer soils due to
414 litter accumulation, and increased edge effects due to population occurrence in fragmented
415 remnants (Jacquemyn *et al.* 2003; Oostermeijer *et al.* 2003; Maurice *et al.* 2012; this study).
416 Plants of *C. glomerata* have been reported to increase in height in dense cover vegetation
417 (Bachmann *et al.* 2005). In translocation sites, nutrient-poor, open grassland conditions were
418 restored by scraping soil, usually followed by livestock grazing, aiming at being optimal for
419 grassland specialist species (Goret *et al.* 2021). The maternal environment of the transplants (in
420 seed source populations) may thus differ from the post-translocation progeny environment.
421 Strong maternal effects still related to marginal ecological conditions might produce maladapted
422 progenies in restored grasslands (Reckinger *et al.* 2010; Schuler & Orrock 2012).

423

424 **Inbreeding depression at early progeny growth stage**

425 Maternal effects can also be associated with maternal inbreeding levels, as in *Campanula*
426 *rapunculoides*, which were reported to affect seed set and seed germination timing, while
427 inbreeding depression was found at all plant life stages (Vogler *et al.* 1999). Inbreeding
428 depression is an important issue for translocated population viability in the long term, as reduced
429 fitness expressed through reduced plant vigour in inbred progeny may result in lower
430 reproductive success and plant survival (Edmands 2007; Aguilar *et al.* 2019). Despite relatively
431 low homozygosity by loci (HL) values (≤ 0.505), which might be expected for self-incompatible
432 species, we found evidence of inbreeding depression (rather than heterosis) for the post-
433 translocation generation of *C. glomerata*, at very early stages of the life cycle. Indeed, rosette
434 diameter at the beginning of growth was smaller for plants showing the highest homozygosity
435 levels. Similarly, two-month old transplants with higher HL values were also smaller (Van
436 Rossum *et al.* 2022). Also, juvenile mortality was the highest for maternal transplants originating
437 from FON, the seed source population showing on average the highest HL values. The similar
438 fitness values found for the two cross categories for the same range of HL values suggest that
439 inbreeding was the main component involved in fitness reduction. In the case of drift load, we
440 might have expected heterosis, i.e. a higher fitness for seed progeny obtained from crosses
441 between seed source origins than from crosses within seed source origins (Paland & Schmid
442 2003). Drift load might also have been expressed at very early stages of development, e.g.
443 through reduced seed germination ability, while inbreeding through juvenile mortality might
444 have contributed to the purging of deleterious recessive alleles (Byers & Waller 1999; Willi *et al.*
445 *et al.* 2013). Given plant longevity, the high (pre-fragmentation) genetic diversity still kept in the
446 old adult generation, and the early stage of genetic erosion in the offspring generation (Van
447 Rossum *et al.* 2022, 2023a), there might also not have been enough generations yet for
448 accumulation and fixation of deleterious mutations due to drift, and so for drift load to be
449 expressed (Willi *et al.* 2013).

450

451 Because of self-incompatibility, inbreeding depression may be related to biparental inbreeding
452 among relatives: higher HL values for progeny obtained from crosses within than between seed

453 sources suggest biparental inbreeding between transplants that may be full sibs or half sibs.
454 Indeed, given the very small size of some seed source populations (flowering population sizes
455 ranging 7 to 28), and evidence of restricted gene flow among populations (Van Rossum *et al.*
456 2022), the number of potential mates for producing the seeds used for propagating transplants is
457 limited, so that we may expect some relatedness between a number of transplants despite
458 extensive within-population pollen dispersal (Barmantlo *et al.* 2018; Aguilar *et al.* 2019; Millar
459 *et al.* 2021). As parentage analyses revealed short-distance seed and pollen dispersal in the two
460 translocated populations of *C. glomerata* (with most dispersal events within a few meters),
461 leading to spatial genetic structure in the newly established generation at a fine scale (Van
462 Rossum *et al.* 2023a), biparental inbreeding and so inbreeding depression issues might still be
463 expected in the next generations. No effect of inbreeding could be detected at later growth
464 stages, but measures were made in good cultivation conditions (especially watering). More
465 stressful hydric conditions on the field (dry grassland habitats) might negatively impact inbred
466 juvenile survival (Willi *et al.* 2006; Edmands 2007).

467

468 **Conclusion**

469 In conclusion, our findings highlighted that maternal transplant origin, especially when seed
470 sources consisted of small fragmented remnants, might possibly have a selective value on fitness
471 at seedling establishment in the post-translocation generations. How maternal effects and
472 inbreeding depression might persist and affect global genetic diversity patterns in the long term
473 might be investigated by comparing the genetic composition of seed progeny with the newly
474 established generation once it has reached the adult stage. Moreover, whether further admixture
475 in the next generations might buffer maternal and inbreeding effects or on the contrary lead to
476 outbreeding depression (Edmands 2007; Barmantlo *et al.* 2018), certainly merits to be
477 investigated, e.g. by comparing fitness of seed progenies identified for their parental seed source
478 origins from successive generations.

479

480 **ACKNOWLEDGEMENTS**

481 This study was funded by the European Union LIFE+ Nature and Biodiversity Program (project
482 no. LIFE11 NAT/BE/001060). We thank the Département de la Nature et des Forêts (Service
483 Public de Wallonie) and Natagora for giving access to the study sites and authorization to collect
484 plant material, S. Godefroid for flower counting of the transplants; A. Van de Vyver, F.
485 Vandeloock, H. Lequeux, M. Du Bois, M. Laureys, G. Mager, J. Smyers, E. Van der Straeten, J.
486 Van Eeckhoudt, G. Van Manen, and M. Verswyvel for help with germination and greenhouse
487 experiments; D. Byers and three anonymous reviewers for constructive comments on the
488 manuscript.

489

490 **SUPPORTING INFORMATION**

491 Additional supporting information may be found online in the Supporting information section at
492 the end of the article.

493

494 **REFERENCES**

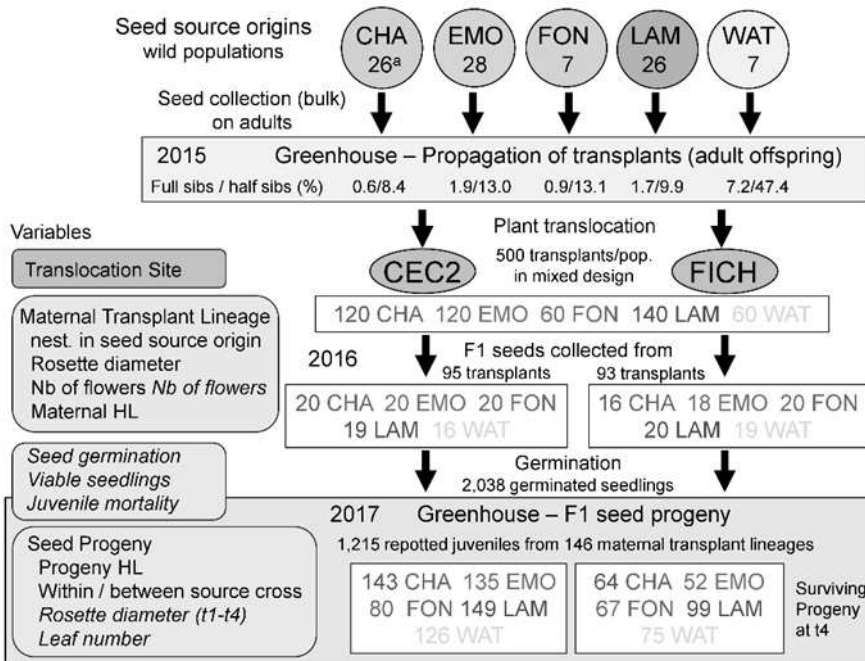
- 495 Aguilar R., Cristóbal-Pérez E.J., Balvino-Olvera F.J., de Jesús Aguilar-Aguilar M., Aguirre-
496 Acosta N., Ashworth L., *et al.* (2019) Habitat fragmentation reduces plant progeny quality: a
497 global synthesis. *Ecology Letters*, **22**, 1163–1173. <https://doi.org/10.1111/ele.13272>
498 Albrecht M., Duelli P., Müller C., Kleijn D., Schmid B. (2007) The Swiss agri-environment
499 scheme enhances pollinator diversity and plant reproductive success in nearby intensively
500 managed farmland. *Journal of Applied Ecology*, **44**, 813–822. <https://doi.org/10.1111/j.1365-2664.2007.01306.x>
501

- 502 Aparicio J.M., Ortego J., Cordero P.J. (2006) What should we weigh to estimate heterozygosity,
 503 alleles or loci? *Molecular Ecology*, **15**, 4659–4665. [https://doi.org/10.1111/j.1365-](https://doi.org/10.1111/j.1365-294X.2006.03111.x)
 504 [294X.2006.03111.x](https://doi.org/10.1111/j.1365-294X.2006.03111.x)
- 505 Bachmann U., Hensen I. (2006) Are population sizes of *Campanula glomerata* on the decline
 506 following the abandonment of traditional land-use practices? *Feddes Repertorium*, **117**, 164–
 507 171. <https://doi.org/10.1002/fedr.200511086>
- 508 Bachmann U., Hensen I. (2007) Is declining *Campanula glomerata* threatened by genetic
 509 factors? *Plant Species Biology*, **22**, 1–10. <https://doi.org/10.1111/j.1442-1984.2007.00170.x>
- 510 Bachmann U., Hensen I., Partzsch M. (2005) Is *Campanula glomerata* threatened by
 511 competition of expanding grasses? *Plant Ecology*, **180**, 257–265.
 512 <https://doi.org/10.1007/s11258-005-3788-0>
- 513 Barmantlo S.E., Meirmans P.G., Luijten S.H., Triest L., Oostermeijer J.G.B. (2018) Outbreeding
 514 depression and breeding system evolution in small, remnant populations of *Primula vulgaris*:
 515 consequences for genetic rescue. *Conservation Genetics*, **19**, 545–554.
 516 <https://doi.org/10.1007/s10592-017-1031-x>
- 517 Bell D.A., Robinson Z.L., Funk W.C., Sarah W., Fitzpatrick S.W., Allendorf F.W., Tallmon
 518 D.A., Whiteley A.R. (2019) The exciting potential and remaining uncertainties of genetic
 519 rescue. *Trends in Ecology & Evolution*, **34**, 1070–1079.
 520 <https://doi.org/10.1016/j.tree.2019.06.006>
- 521 Byers D.L., Waller D.M. (1999) Do plant populations purge their genetic load? effects of
 522 population size and mating history on inbreeding depression. *Annual Review of Ecology and*
 523 *Systematics*, **30**, 479–513. <https://doi.org/10.1146/annurev.ecolsys.30.1.479>
- 524 Coulon A. (2010) GENHET: an easy-to-use R function to estimate individual heterozygosity.
 525 *Molecular Ecology Resources*, **10**, 167–169. [https://doi.org/10.1111/j.1755-](https://doi.org/10.1111/j.1755-0998.2009.02731.x)
 526 [0998.2009.02731.x](https://doi.org/10.1111/j.1755-0998.2009.02731.x)
- 527 Denisow B., Wrzesień M. (2015) The habitat effect on the diversity of pollen resources in
 528 several *Campanula* spp. – an implication for pollinator conservation. *Journal of Apicultural*
 529 *Research*, **54**, 62–71. <https://doi.org/10.1080/00218839.2015.1030243>
- 530 Dillon R., Coates D., Standish R., Monks L., Waycott M. (2023) Assessing plant translocation
 531 success: common metrics mask high levels of inbreeding in a recently established *Banksia*
 532 *brownii* (Proteaceae) population. *Australian Journal of Botany*, **71**, 79–92.
 533 <https://doi.org/10.1071/BT22071>
- 534 Doyle C.A.T., Yap J.-Y.S., Bragg J., Rossetto M., Orme A., Ooi M.J.K. (2023) Reproductive
 535 characteristics, population genetics, and pairwise kinship inform strategic recovery of a plant
 536 species in a fragmented landscape. *Conservation Science and Practice*, **5**, e12910.
 537 <https://doi.org/10.1111/csp2.12910>
- 538 Edmands S. (2007) Between a rock and a hard place: evaluating the relative risks of inbreeding
 539 and outbreeding for conservation and management. *Molecular Ecology*, **16**, 463–475.
 540 <https://doi.org/10.1111/j.1365-294X.2006.03148.x>
- 541 Fenner M., Thompson K. (2005) *The ecology of seeds*. Cambridge University Press, New York.
- 542 Fenu G., Calderisi G., Boršić I., Bou Dagher Kharrat M., García Fernández A., Kahale R.,
 543 Panitsa M., Cogoni D. (2023) Translocations of threatened plants in the Mediterranean
 544 Basin: current status and future directions. *Plant Ecology*, **224**, 762–755.
 545 <https://doi.org/10.1007/s11258-023-01303-7>
- 546 Gadella T.W.J. (1964) Cytotaxonomic studies in the genus *Campanula*. *Wendtia*, **11**, 1–104.
- 547 Galloway L.F. (2001) The effect of maternal and paternal environments on seed characters in the
 548 herbaceous plant *Campanula americana* (Campanulaceae). *American Journal of Botany*, **88**,
 549 832–840. <https://doi.org/10.2307/2657035>
- 550 Galloway L.F. (2005) Maternal effects provide phenotypic adaptation to local environmental
 551 conditions. *New Phytologist*, **166**, 93–100. <https://doi.org/10.1111/j.1469-8137.2004.01314.x>

- 552 Gargiulo R., Worswick G., Arnold C., Pike L.J., Cowan R.S., Hardwick K.A., Chapman T., Fay
 553 M.F. (2019) Conservation of the threatened species, *Pulsatilla vulgaris* Mill. (Pasqueflower),
 554 is aided by reproductive system and polyploidy. *Journal of Heredity*, **110**, 618–628.
 555 <https://doi.org/10.1093/jhered/esz035>
- 556 Godefroid S., Le Pajolec S., Van Rossum F. (2016) Pre-translocation considerations in rare plant
 557 reintroductions: implications for designing protocols. *Plant Ecology*, **217**, 1693–182.
 558 <https://doi.org/10.1007/s11258-015-0526-0>
- 559 Goret T., Janssens X., Godefroid S. (2021) A decision-making tool for restoring lowland
 560 grasslands in Europe. *Journal for Nature Conservation*, **63**, 126046.
 561 <https://doi.org/10.1016/j.jnc.2021.126046>
- 562 Hamilton J.A., Royauté R., Wright J.W., Hodgskiss P., Ledig F.T. (2017) Genetic conservation
 563 and management of the California endemic, Torrey pine (*Pinus torreyana* Parry): Implications
 564 of genetic rescue in a genetically depauperate species. *Ecology and Evolution*, **7**, 7370–7381.
 565 <https://doi.org/10.1002/ece3.3306>
- 566 Hill M.O., Preston C.D., Roy D.B. (2004) *PLANTATT - Attributes of British and Irish plants: status, size, life history, geography and habitats*. NERC Centre for Ecology and Hydrology,
 567 Huntingdon, UK.
- 569 Jacquemyn H., Van Rossum F., Brys R., Endels P., Hermy M., Triest L., De Blust G. (2003)
 570 Effects of agricultural land use and fragmentation on genetics, demography and population
 571 persistence of the rare *Primula vulgaris*, and implications for conservation. *Belgian Journal*
 572 *of Botany*, **136**, 5–22. <https://doi.org/10.2307/20794510>
- 573 Klotz S., Kühn I., Durka W. (2002) BIOLFLOR - Eine Datenbank zu biologisch-ökologischen
 574 Merkmalen der Gefäßpflanzen in Deutschland. *Schriftenreihe für Vegetationskunde*, **38**, 1-
 575 334.
- 576 Koutsovoulou K., Daws M.I., Thanos C.A. (2014) Campanulaceae: a family with small seeds
 577 that require light for germination. *Annals of Botany*, **113**, 135–143.
 578 <https://doi.org/10.1093/aob/mct250>
- 579 [Dataset] Le Pajolec S., Vandeloock F., Van Rossum F., Godefroid S. (2021) Fitness measures of
 580 seed progeny from seven natural populations of *Campanula glomerata*. *Zenodo Digital*
 581 *Repository*. <https://doi.org/10.5281/zenodo.5676272>
- 582 Lofflin D.L., Kephart S.R. (2005) Outbreeding, seedling establishment, and maladaptation in
 583 natural and reintroduced populations of rare and common *Silene douglasii*
 584 (Caryophyllaceae). *American Journal of Botany*, **92**, 1691–1700.
 585 <https://doi.org/10.3732/ajb.92.10.1691>
- 586 Maurice T., Colling G., Muller S., Matthies D. (2012) Habitat characteristics, stage structure and
 587 reproduction of colline and montane populations of the threatened species *Arnica montana*.
 588 *Plant Ecology*, **21**, 831–842. <https://doi.org/10.1007/s11258-012-0045-1>
- 589 McCullagh P., Nelder J. (1989) *Generalized Linear Models*. Second edition. Chapman and Hall,
 590 London, UK.
- 591 Millar M.A., Coates D.J., Byrne M., Krauss S.L., Jonson J., Hopper S.D. (2021) Evaluating
 592 restoration outcomes through assessment of pollen dispersal, mating system, and genetic
 593 diversity. *Restoration Ecology*, **29**, e13335. <https://doi.org/10.1111/rec.13335>
- 594 Mitchell R.J., Flanagan R.J., Brown B.J., Waser N.M., Karron J.D. (2009) New frontiers in
 595 competition for pollination, *Annals of Botany*, **103**, 1403–1413.
 596 <https://doi.org/10.1093/aob/mcp062>
- 597 Monks L., Standish R., McArthur S., Dillon R., Byrne M., Coates D. (2021) Genetic and mating
 598 system assessment of translocation success of the long-lived perennial shrub *Lambertia*
 599 *orbifolia* (Proteaceae). *Restoration Ecology*, **29**, e13369. <https://doi.org/10.1111/rec.13369>

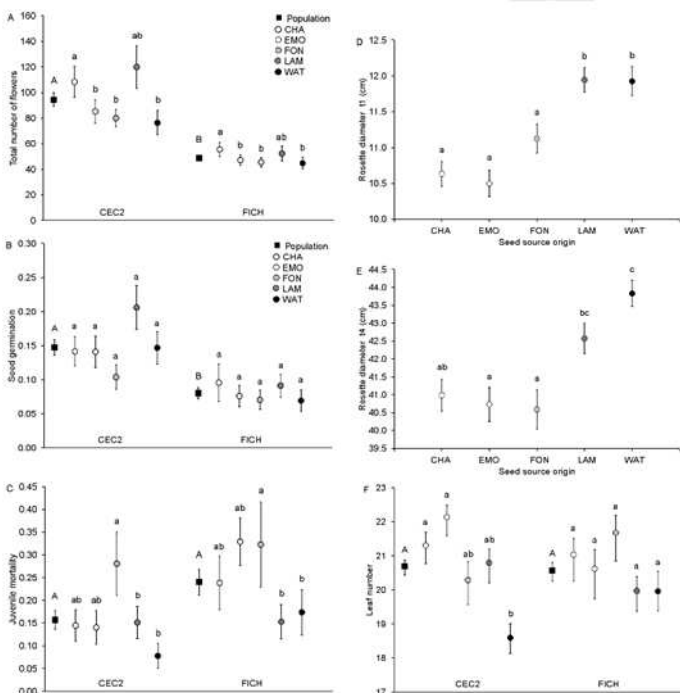
- 600 Nicotra A.B., Beever E.A., Robertson A.L., Hofmann G.E., O'Leary J. (2015) Assessing the
601 components of adaptive capacity to improve conservation and management efforts under
602 global change. *Conservation Biology*, **29**, 1268–1278. <https://doi.org/10.1111/cobi.12522>
- 603 Oostermeijer J.G.B., Luijten S.H., den Nijs J.C.M. (2003) Integrating demographic and genetic
604 approaches in plant conservation. *Biological Conservation*, **113**, 389–398.
605 [https://doi.org/10.1016/S0006-3207\(03\)00127-7](https://doi.org/10.1016/S0006-3207(03)00127-7)
- 606 Paland S., Schmid B. (2003) Population size and the nature of genetic load in *Gentianella*
607 *germanica*. *Evolution*, **57**, 2242–2251. <https://doi.org/10.1111/j.0014-3820.2003.tb00236.x>
- 608 Pritchard J.K., Stephens M., Donnelly P. (2000) Inference of population structure using
609 multilocus genotype data. *Genetics*, **155**, 949–959.
- 610 Reckinger C., Colling G., Matthies D. (2010) Restoring populations of the endangered plant
611 *Scorzonera humilis*: influence of site conditions, seed source, and plant stage. *Restoration*
612 *Ecology*, **18**, 904–913. <https://doi.org/10.1111/j.1526-100X.2009.00522.x>
- 613 Roach, D.A., Wulff, R.D. (1987) Maternal effects in plants. *Annual Review of Ecology and*
614 *Systematics*, **18**, 209–235. <http://www.jstor.org/stable/2097131>
- 615 Rodger Y.S., Dillon R., Monro K., Pavlova A., Coates D.J., Byrne M., Sunnucks P. (2023)
616 Benefits of outcrossing and their implications for genetic management of an endangered
617 species with mixed-mating system. *Restoration Ecology*, **32**, e14057.
618 <https://doi.org/10.1111/rec.14057>
- 619 Schuler M.S., Orrock, J.L. (2012) The maladaptive significance of maternal effects for plants in
620 anthropogenically modified environments. *Evolutionary Ecology*, **26**, 475–481.
621 <https://doi.org/10.1007/s10682-011-9499-1>
- 622 Van Rossum F. (2023) Sibship and parentage reconstruction as genetic tool for designing and
623 monitoring plant translocations. *Restoration Ecology*, **31**, e13726.
624 <https://doi.org/10.1111/rec.13726>
- 625 [Dataset] Van Rossum F., Godé C. (2021) Individual multilocus genotypes of adults and seed
626 progeny from eight natural populations of *Campanula glomerata*. *Zenodo Digital Repository*.
627 <https://doi.org/10.5281/zenodo.5676286>
- 628 Van Rossum F., Godé C. (2022) Development of highly polymorphic microsatellite markers for
629 the declining *Campanula glomerata* (Campanulaceae). *Molecular Biology Reports*, **49**, 805–
630 810. <https://doi.org/10.1007/s11033-021-06839-3>
- 631 [Dataset] Van Rossum F., Godé C. (2023) Individual multilocus genotypes of F1 seed progeny
632 and recruits from two translocated populations of *Campanula glomerata*. *Zenodo Digital*
633 *Repository*. <https://doi.org/10.5281/zenodo.7954368>
- 634 [Dataset] Van Rossum F., Godefroid S., Le Pajolec S. (2023b) Fitness measures of maternal
635 transplants and their seed progeny from two translocated populations of *Campanula*
636 *glomerata* in southern Belgium. *Zenodo Digital Repository*.
637 <https://doi.org/10.5281/zenodo.8217417>
- 638 Van Rossum F., Hardy O.J. (2022) Guidelines for genetic monitoring of translocated plant
639 populations. *Conservation Biology*, **36**, e13670. <https://doi.org/10.1111/cobi.13670>
- 640 Van Rossum F., Hardy O.J., Le Pajolec S., Raspé O. (2020) Genetic monitoring of translocated
641 plant populations in practice. *Molecular Ecology*, **29**, 4040–4058.
642 <https://doi.org/10.1111/mec.15550>
- 643 Van Rossum F., Le Pajolec S. (2021) Mixing gene pools to prevent inbreeding issues in
644 translocated populations of clonal species. *Molecular Ecology*, **30**, 2756–2771.
645 <https://doi.org/10.1111/mec.15930>
- 646 Van Rossum F., Le Pajolec S., Godé C. (2023a) Assessing spatial mating patterns in translocated
647 populations of *Campanula glomerata*. *Global Ecology and Conservation*, **46**, e02548.
648 <https://doi.org/10.1016/j.gecco.2023.e02548>

- 649 Van Rossum F., Le Pajolec S., Raspé O., Godé C. (2022) Assessing population genetic status for
650 designing plant translocations. *Frontiers in Conservation Science*, **3**, 829332.
651 <https://doi.org/10.3389/fcosc.2022.829332>
- 652 Vergeer P., Sonderen E., Ouborg N.J. (2004) Introduction strategies put to the test: local
653 adaptation versus heterosis. *Conservation Biology*, **18**, 812–821.
654 <https://doi.org/10.1111/j.1523-1739.2004.00562.x>
- 655 Vogler D.W., Filmore K., Stephenson A.G. (1999) Inbreeding depression in *Campanula*
656 *rapunculoides* L. I. A comparison of inbreeding depression in plants derived from strong and
657 weak self-incompatibility phenotypes. *Journal of Evolutionary Biology*, **12**, 483–494.
658 <https://doi.org/10.1046/j.1420-9101.1999.00046.x>
- 659 Willi Y., Griffin P., Van Buskirk J. (2013) Drift load in populations of small size and low
660 density. *Heredity*, **110**, 296–302. <https://doi.org/10.1038/hdy.2012.86>
- 661 Willi Y., Van Buskirk J., Fischer M. (2005) A threefold genetic Allee effect: population size
662 affects cross-compatibility, inbreeding depression and drift load in the self-incompatible
663 *Ranunculus reptans*. *Genetics*, **169**, 2255–2265. <https://doi.org/10.1534/genetics.104.034553>
- 664 Willi Y., Van Buskirk J., Hoffmann A.A. (2006) Limits to the adaptive potential of small
665 populations. *Annual Review of Ecology, Evolution, and Systematics*, **37**, 433–458.
666 <https://doi.org/10.1146/annurev.ecolsys.37.091305.110145>
- 667 Zavodna M., Abdelkrim J., Pellissier V., Machon N. (2015) A long-term genetic study reveals
668 complex population dynamics of multiple-source plant reintroductions. *Biological*
669 *Conservation*, **192**, 1–9. <https://doi.org/10.1016/j.biocon.2015.08.025>
- 670 Zimmer H., Auld T., Cuneo P., Offord C., Commander L. (2019) Conservation translocation -
671 An increasingly viable option for managing threatened plant species. *Australian Journal of*
672 *Botany*, **67**, 501–509. <https://doi.org/10.1071/BT19083>
- 673



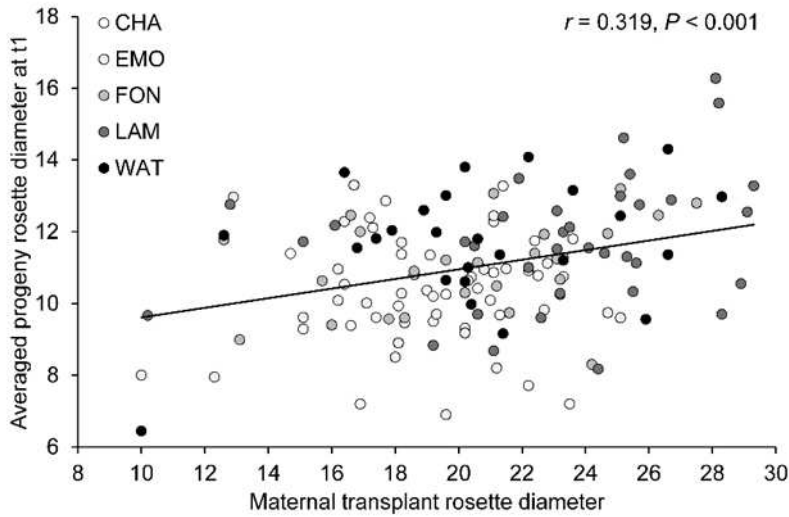
674
675
676
677
678
679
680

Fig. 1. Summary of sampling and translocation design and of the greenhouse experiment on F1 seed progeny, with the tested response (in italics) and predictor variables in this study (number of flowers of maternal transplants was both a response and a predictor variable; see Table S1). Percentage of full sibs and half sibs in transplants for each seed source population from Van Rossum *et al.* (2022). ^aNumber of flowering adults.



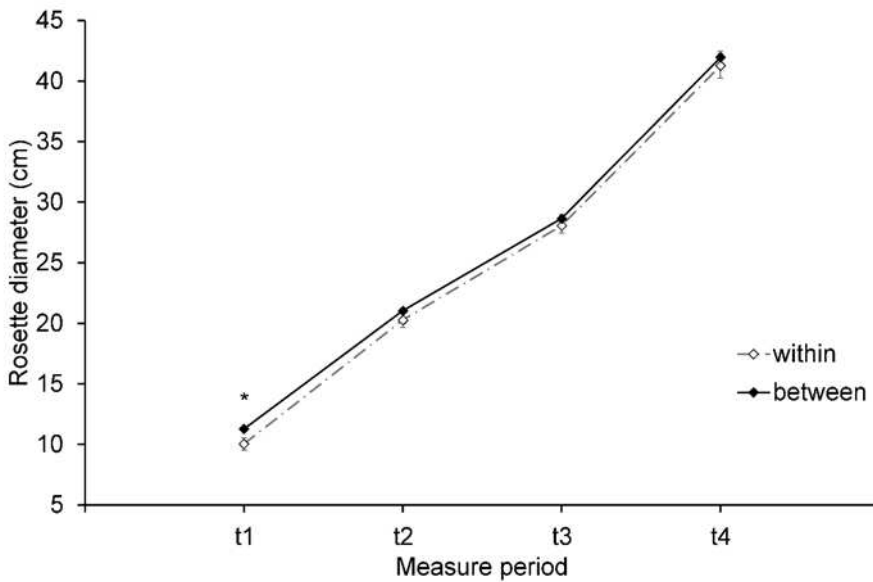
681
682
683
684
685
686
687
688

Fig. 2. Fitness variables (mean value \pm SE) of the maternal transplants (a-b) or seed progeny (c-f) according to translocation site (CEC2, FICH), and maternal seed source origin (CHA, EMO, FON, LAM and WAT): (a) total number of flowers; (b) seed germination; (c) juvenile mortality; (d) rosette diameter (cm) at t1; (e) rosette diameter (cm) at t4; and (f) leaf number. Different symbols indicate significant ($P < 0.05$) differences between sites (capital letters) based on ANCOVA (F tests) or Generalized Linear Model (Wald tests), and among source origins (small letters) based on Tukey tests.



689
690
691
692

Fig. 3. Rosette diameter (mean value per maternal transplant in cm) of seed progeny at t1 as a function of rosette diameter of maternal transplants (in cm). *r*, Pearson's correlation coefficient.



693
694
695
696
697

Fig. 4. Rosette diameter (mean \pm SE) of seed progeny according to cross category (within/between seed source origins). * $P < 0.05$ according to Mann-Whitney *U* test between cross categories.