

1 *Pseudostaurosira ellipticolanceolata*, a new araphid diatom (Bacillariophyta) from Flanders,  
2 Belgium

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4 Coralie André<sup>1,5</sup>, Koen Sabbe<sup>2,6</sup> & Bart Van de Vijver<sup>3,4,7\*</sup>

5  
6 <sup>1</sup>Ghent University, Department of Geology, Krijgslaan 281-S8, 9000 Ghent, Belgium

7 <sup>2</sup>Ghent University, Protistology and Aquatic Ecology Lab, Krijgslaan 281-S8, 9000 Ghent,  
8 Belgium

9 <sup>3</sup>Meise Botanic Garden, Research Department, Nieuwelaan 38, 1860 Meise, Belgium;

10 <sup>4</sup>University of Antwerp, Department of Biology – ECOSPHERE, Universiteitsplein 1, 2610  
11 Wilrijk, Belgium

12  
13 <sup>5</sup>[coralie.andre@ugent.be](mailto:coralie.andre@ugent.be), <https://orcid.org/0000-0001-5933-7773>

14 <sup>6</sup>[koen.sabbe@ugent.be](mailto:koen.sabbe@ugent.be), <https://orcid.org/0000-0001-5163-5581>

15 <sup>7</sup>[bart.vandevijver@plantentuinmeise.be](mailto:bart.vandevijver@plantentuinmeise.be), <https://orcid.org/0000-0002-6244-1886>

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17 \*corresponding author

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19 During the analysis of a sediment core collected in Aardenburg (Flanders, Belgium), species of a  
20 species of *Pseudostaurosira* that could not be identified using the currently available literature  
21 was observed. The genus *Pseudostaurosira* D.M. Williams & Round (1987: 276) contains mostly  
22 species with an elliptic, linear to linear-lanceolate valve outline, possessing uniseriate striae  
23 composed of one to only a few small, circular areolae, occluded by usually branched volae, a  
24 broad sternum, small, reduced apical pore fields, that occasionally are absent, and the absence of  
25 rimoportulae (Williams & Round 1987, Cejudo-Figueras *et al.* 2011). Recent taxonomic  
26 advances in the genus *Pseudostaurosira* (Cejudo-Figueiras *et al.* 2011, Garcia *et al.* 2021, Grana  
27 *et al.* 2018, Li *et al.* 2018, Morales *et al.* 2015, 2019, 2021) markedly increased our knowledge  
28 of this group. Morales *et al.* (2021) illustrated and discussed many new species of  
29 *Pseudostaurosira*, providing a detailed overview of a large number of species in this genus. To  
30 date, however, the morphological delineation of the genus *Pseudostaurosira* is still debated (see  
31 e.g. Marquardt *et al.* 2021), and its exact molecular phylogenetic status and position are as yet  
32 unresolved, with the genus being potentially paraphyletic (Li *et al.* 2018). We here adopted the  
33 above-mentioned list of features as being diagnostic of the genus *Pseudostaurosira*, and have  
34 therefore decided to assign the new species to this genus.

35 Despite extensive comparison with morphologically similar *Pseudostaurosira* species  
36 worldwide, the unknown species in the Aardenburg sediment core could not be identified and  
37 therefore most likely represents a new species that is described here based on light (LM) and  
38 scanning electron microscopy (SEM) observations: *Pseudostaurosira ellipticolanceolata* André,  
39 Sabbe & Van de Vijver, *sp. nov.* As we are dealing with subfossil material, the valves are partly  
40 eroded obscuring some details such as spine shape and areola occlusions. Nevertheless, the  
41 combination of observed features enabled to separate the new species from all previously  
42 described *Pseudostaurosira* species. The associated diatom flora is used to provide an ecological  
43 profile of the new species.

44

#### 45 **New species description**

46 ***Pseudostaurosira ellipticolanceolata* André, Sabbe & Van de Vijver, *sp. nov.*** (Figs 1–21 LM,  
47 22–28 SEM)

48 Valves lanceolate to linear-lanceolate in larger specimens, becoming elliptic-lanceolate to elliptic  
49 in smallest valves (Figs 1–21). Valve margins parallel in longer valves to distinctly convex in the  
50 smaller range of the cell diminution series. Apices very weakly protracted (Figs 1–3),  
51 subrostrate, to non-protracted, broadly rounded in smaller specimens. Valve dimensions (n=40):  
52 valve length 4–20 µm, valve width 3.5–4.0 µm. Sternum broad, lanceolate. Striae uniseriate,  
53 parallel to weakly radiate throughout the entire valve length, 16–18 in 10 µm, composed of 1–2,  
54 rarely 3 areolae, on valve face (Figs 22, 25), and 1–2 areolae on valve mantle. Areolae at valve  
55 face/mantle junction markedly larger, becoming smaller towards sternum (Figs 22, 24). Mantle  
56 areolae separated from those on valve face by short linking spines (Figs 22, 25). Interdigitating  
57 spines originating from vimines, spatulate with broad tip (but probably broken and eroded) (Fig.  
58 23, white arrows). Areola occlusions eroded, vestiges partly visible in some areolae, suggesting  
59 volate occlusion type (Fig. 28, black arrows). Apical pore field reduced to a handful of very  
60 small pores (Figs 27, 28, white arrows). Internally, sternum flat with raised virgae between striae  
61 (Fig. 26). Rimoportula absent. Girdle structure not observed due to severe erosion of the  
62 frustules.

63

64 **Type:**—BELGIUM. Aardenburg sediment core, sample 85, coll. date 1<sup>st</sup> of March 2019, leg. C.  
65 André (holotype slide BR-XXXX= Fig. XXX, isotype slide XXXX, University of Antwerp,  
66 Belgium). Sample 85 is part of a silty clay layer.

67 **Phycobank registration:** —<https://phycobank.org/XXXX>

68  
69 **Ecology & associated diatom flora:**—*P. ellipticolanceolata* has a relative abundance of almost  
70 10% in the type sample. The sample is part of sediment core and dates back to the Holocene. The  
71 first indications show a possible age between the 4<sup>th</sup> and 10<sup>th</sup> century AD (André, unpubl. res.).  
72 Other species with a relative abundance greater than 3% include (in decreasing abundance)  
73 *Epithemia adnata* (Kützing 1833: 544) Brébisson (1838: 16), *Staurosira venter* (Ehrenberg 1854:  
74 13, pl. XIV) Cleve & J.D.Möller (1879: no. 242) sensu Lange-Bertalot *et al.* (2017), *Eunotia minor*  
75 (Kützing 1844: 39) Grunow (in Van Heurck 1881: pl. 33, figs 20, 21), *Cymatosira belgica* Grunow  
76 (in Van Heurck 1881: pl. 45: figs 38–41), *Navicula cincta* (Ehrenberg 1854: pl. 10/2: fig. 6a-e)  
77 Ralfs (in Pritchard 1861: 901), *Epithemia turgida* (Ehrenberg 1832: 80) Kützing (1844: 34),  
78 *Nanofrustulum sopotensis* (Witkowski & Lange-Bertalot 1993: 67) E.Morales *et al.* (in Morales *et*  
79 *al.* 2019: 275), *Paralia sulcata* (Ehrenberg 1838: 170) Cleve (1873: 7) and *Gomphonema*  
80 *exilissimum* (Grunow in Van Heurck 1880: pl. 25: fig. 12) Lange-Bertalot & E.Reichardt (in  
81 Lange-Bertalot & Metzeltin 1996: 70). This diatom flora points to a brackish, eutrophic  
82 environment (Lange-Bertalot *et al.* 2017). Both *Epithemia* species are typically epiphytic and may  
83 point to the presence of submerged vegetation, but are also salinity-tolerant and often occur in  
84 aquatic conditions with higher electrolyte contents (Lange-Bertalot *et al.* 2017). McQuoid &  
85 Nordberg (2003) observed *Paralia sulcata* to be dominant in well-mixed upwelling zones with  
86 high salinities and high phosphorus levels. *Paralia* and *Cymatosira* often co-occur. Both taxa are  
87 considered marine/marine-brackish tycho planktonic and observed typically in tidal inlets and large  
88 tidal channels (Vos & de Wolf, 1988). The presence of *Eunotia minor* is more difficult to explain  
89 but most likely the result of the influx of foreign material.

90  
91 **Taxonomic comments:**—Morales *et al.* (2021) discuss a large number of *Pseudostaurosira*  
92 species that are known worldwide when they described several new species from Bolivia. Based  
93 on valve outline and valve dimensions, the new *P. ellipticolanceolata* shows some similarity  
94 with the group of *Pseudostaurosira subsalina* (Hustedt) E.A.Morales (2005: 115) and *P.*  
95 *polonica* (M.Witak & Lange-Bertalot in Witkowski *et al.* 1995: 736) E.A.Morales &  
96 M.B.Edlund (2003: 235). *Pseudostaurosira occulta* E.A.Morales, C.E.Wetzel & Ector (2021: 40)  
97 shows comparable valve dimensions (length 7–35 µm, width 3.5–4.0 µm) and, at least in the  
98 longer range of the size diminution series, a comparable valve outline with almost parallel  
99 margins. However, differences can be noted in the shape of the apices (squarish in *P. occulta*,  
100 more broadly rounded in *P. ellipticolanceolata*), stria density (14–16 in 10 µm in *P. occulta*  
101 versus 16–18 in 10 µm in *P. ellipticolanceolata*) and stria structure. In *P. occulta*, striae are  
102 usually composed of 3–4 areolae whereas striae with 3 areolae are rather rare in *P.*  
103 *ellipticolanceolata*. Due to the high degree of erosion, it was not possible to comment on the  
104 structure and shape of the apical pore field (covered by an external flap in *P. occulta*) and the  
105 linking spines. *Pseudostaurosira subsalina* has broader valves (valve width 4.0–5.5 µm), a lower  
106 stria density (13–14 in 10 µm), a higher number of areolae per stria, a larger apical pore field  
107 (compared to the reduced pore field in *P. ellipticolanceolata*), and a broader transition step  
108 between valve face and mantle, excluding possible conspecificity (Cejudo-Figueiras *et al.* 2011).  
109 *Pseudostaurosira zolitschkae* M.L.García *et al.* (2021: 265) differs in having acutely protracted

110 valve apices and a much lower stria density (11–14 in  $\mu\text{m}$ ) (García *et al.* 2021). Other taxa in this  
111 group differ by the shape of the areolae (*P. polonica* has only one, transapically elongated areola  
112 per striae), valve dimensions or valve outline. *Pseudostaurosira oliveraiana* Grana *et al.* (2018:  
113 63) has longer valves (19–39  $\mu\text{m}$ ) and typically capitate to rostrate apices throughout its entire  
114 cell diminution series (Grana *et al.* 2018, figs 2–15). Smaller taxa such as *P. alvareziae* Cejudo-  
115 Figueiras, E.A.Morales & Ector (2011: 69) and *P. oblonga* E.A.Morales, C.E.Wetzel & Ector  
116 (2021: 41) have a more elliptic to elliptic-lanceolate valve outline with strictly convex margins,  
117 whereas elongated, more linear-lanceolate valves are so far not observed, in contrast to *P.*  
118 *ellipticolanceolata* where longer valves usually have parallel margins.

119

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125

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214

215 **Figure captions**

216 Figures 1–35. *Pseudostaurosira ellipticolanceolata* André, Sabbe & Van de Vijver, sp. nov. LM  
217 and SEM pictures taken from the holotype material (BR-XXX, Aardenburg sediment core,  
218 depth 85 cm, Belgium). Figs 1–21. LM views of several specimens arranged in decreasing  
219 size. Fig. 22. SEM external view of an entire valve. Fig. 23. SEM view of two valves with  
220 interdigitating spines (white arrows). Fig. 24. SEM internal view of an entire valve,  
221 connected to another. Fig. 25. SEM external view of an entire valve. Fig. 26. SEM internal  
222 view of an entire valve. Note the raised virgae bordering the flat sternum. Fig. 27. SEM  
223 internal detail of the valve apex. The white arrow indicates the apical pore field. Fig. 28.  
224 SEM external detail of the valve apex. The white arrows indicate the eroded apical pore  
225 field. The black arrows indicate the presence of vestiges of the areola coverings. Scale bars  
226 represent 10 µm except for Figs 23, 25 & 27 where scale bar = 5 µm and Fig. 28 where  
227 scale bar = 1 µm.  
228

